

## INCIDENCE OF PAPULASPORA BYSSINA (BROWN PLASTER MOULD) ON CASING MIXTURE AND COMPOST OF WHITE BUTTON MUSHROOM (AGARICUS BISPORUS)

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#### **KEYWORDS**

Agaricus bisporus Papulaspora byssina Biological efficiency

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#### **INTRODUCTION**

#### ABSTRACT

During cultivation of white button mushroom (*Agaricus bisporus*), at Sam Higginbottom Institute of Agriculture, Technology and Sciences, Allahabad heavy infestation of *Papulaspora byssina* was observed on casing mixture and compost during 2012 and 2013. Therefore, an investigation was carried out to manage the brown plaster mould by modifying the casing materials used. Altogether seven treatments were taken up for this study. These treatments were replicated seven times and data collected was analyzed using CRD. *P. byssina* infected compost at very early stage after casing in the Allahabad agro-climatic conditions. The treatment combination of spent mushroom compost with sand and lime (4:1:1) showed highest incidence of *P. byssina* (83.34%) which reduced the yield (.250kg/bag) and biological efficiency (4.46%) as compared with other treatments. While the treatments farm yard manure with garden loam soil (2:1) and farm yard manure with garden loam soil and sand (2:1:1) showed lowest incidence of *P. byssina* (33.00%) which gave the better yield (.750kg/bag), (.875kg/bag) and biological efficiency (18.05%), (23.02%) respectively.

Mushroom cultivation is a potential biotechnological process wherein the waste plant materials or negative value crop residues can be converted into valuable food. Protein conversion efficiency and productivity of mushrooms per unit land area and time is far superior than plant and animal sources. In 2007, the production of edible mushrooms in Japan was estimated to be 4, 23, 224 tones and it is expected that this amount will increase in the future due to market demand. (Shitole, 2014). Recently, mushroom cultivation in India has witnessed a tremendous growth with respect to the type of mushrooms and their productivity. Mushrooms are considerd as valuable health food since they are known for rich proteinacious food, it consists of about 75% proteins and are low in calories, fat, fatty acids, vitamins and minerals. (Sharma, 2013). The mushroom market is growing continuously mainly due to increasing interest in their culinary, nutritional and health benefits. On recognizing the importance of mushroom which can be an eco-friendly alternative for agro-waste recycling, capability to provide better nutrition for the vast vegetarian population, employment generation and a good source of income (AICRP, 2008).

*Agaricus bisporus* is cultivated in more than 70 countries in the world. The annual world production of button mushroom has reached 3.5 million tones and that of all types of mushrooms is estimated to be over 20 million tones. It contributes about 90 per cent of total country's production as against its global share of about 40 per cent (**Mehta et al.**, 2011). Our country has registered twenty-fold increase in production of mushrooms in the last four decades, even

though, button mushroom continues to occupy a prominent place and contributes about 85% of the total mushroom production of the country. Mushroom vegetative growth was done in compost or culture bed. In different countries, different materials are used for compost production using different organic wastes which could be useful in increasing the production. Reproductive growth and fruit body production occurred in the layer named casing layer, in which mushroom growth and yield were increased. (Ebadi, 2012). . Many pathogens and pests affect this monoculture. The most important pathogens of A. bisporus are green moulds (Trichoderma sp.), Papulaspora byssina, Coprinus sp., Rhizoctonia sp., Lecanicillium fungicola etc. The number and composition of microorganisms which accompany mushroom cultivation depends on the healthiness of the compost, casing and A. bisporus spawns. (Amra et al., 2008). Papulaspora byssina (Brown Plaster mould) is also reported to cause 90-92% yield loss in A. bisporus. This mould has also been reported to cause complete crop failure in oyster mushroom in Kasuali, HP (Anonymous, 2012). The aim of this study was selection of suitable casing materials using different agricultural wastes which can inhibit the enhanced growth and incidence of P. byssina

#### MATERIALS AND METHODS

The experimental trial was conducted at Mushroom Crop Room in Department of Plant Protection, SHIATS, Allahabad with seven treatment combinations *viz*. Farm Yard Manure (FYM) + Garden loam soil (GLS) (2:1), FYM + GLS + Sand (2:1:1), FYM + GLS + Sawdust (2:1:1), FYM + Sawdust + Sand (2:1:1), FYM + GLS + Waste Tea Leaves (2:1:1), FYM + GLS + Vermi-compost (2:1:1) and Spent Mushroom Compost (SMC) + Sand + Lime (4:1:1). These treatments were replicated seven times and data collected was analyzed using CRD. The maximum temperature and relative humidity during October 2012 to February 2013 were 38.2°C and 97%, respectively while the minimum temperature and relative humidity during this period were 6.2°C and 32%, respectively. Compost was prepared by long method of composting using wheat straw (Mantel et *al.*, 1972).

#### **RESULTS AND DISCUSSION**

During the cultivation of white button mushroom, after 15-17 days of casing some light orange coloured patches appeared on casing soil. After 2-3 days the diameter and frequency of patches increased and spread to compost. In the later stage of infection orange colour liquid was observed (Fig. 1). After 28-30 days of infection, the incidence of Papulaspora byssina was recorded in the range of 34-84% (Table 1). Isolation of fungus was carried out on Potato Dextrose Agar (PDA) medium and petri plates were incubated for 2-3 days at 25°C and a pure culture of fungus was obtained (Fig. 2). Microscopic observation showed the presence of orangish brown mycelium which produced brown coloured many celled spherical, bulbils which were interwoven with network of hyphae, were set free later with the death of the mycelium (Fig. 3). Maximum average yield and biological efficiency (%) (0.875 kg/bag and 23.02, respectively) were recorded from treatment combination of FYM with GLS and sand (2:1:1) (Table 1) and highest incidence of P. byssina (83.34%) was recorded from treatment combination of SMC with sand and lime (4:1:1) (Table 1). This disease has also been reported from India (Munjal and Seth, 1974) causing 90-92% yield loss in A. bisporus. The treatment combination of spent mushroom compost with sand and lime (4:1:1) also reduced the yield (.250kg/bag) and biological efficiency (4.46%) as compared with other treatments. Loss in number and weight of fruit bodies as a result of artificial inoculation of the mould has been found 7.7-53.5% and 3.0-50.7% respectively (Sharma, 1990; Sharma and Vijay, 1993). Comparing the results with C.D value, it was found that FYM+ GLS+ Sand (2:1:1) was significantly superior over all treatments.

The probable reason for such finding may be that the primary inoculum of pathogen already existed in the SMC as it was earlier used as casing mixture. The combination of lime and SMC provided appropriate atmospheric conditions for development of pathogen. Carbon di oxide is released by CaCO<sub>3</sub> may have favoured the growth of *P. byssina*. Perusal of available literature revealed that incidence of *Papulaspora byssina* on the compost of button mushroom was reported from Ludhiana and Pantnagar (AICRP, 2005-06) and on the substrate of milky mushroom from Allahabad (Uttar Pradesh) (Kumar et al., 2013). This fungus now is frequently found at almost all the mushroom farms in India appearing usually during spawn run (Garcha et al., 1987; Kaul et al., 1978; Sharma, 1992).



Figure 1: Papulaspora byssina on compost of Agaricus bisporus



Figure 2: Pure culture of Papulaspora byssina

Table 1: Effect of casing materials on the	vield and biological efficience	v of A. bisporus and incidence of P. b	vssina on compost of A. bisporus

Treatments	Incidence of P. byssina* (%)	Yield*(Kg/bag**)	BiologicalEfficiency (%)
Farm Yard Manure (FYM) + Garden loam soil (GLS) (2:1)	33.00	0.750	18.05
FYM + GLS + Sand (2:1:1)	33.00	0.875	23.02
FYM+ GLS+ Sawdust (2:1:1)	50.00	0.694	16.50
FYM+ Sawdust+ Sand (2:1:1)	50.00	0.684	16.26
FYM+ GLS+ Waste Tea Leaves (2:1:1)	50.00	0.608	12.16
FYM+ GLS+ Vermi-compost (2:1:1)	66.67	0.4775	8.48
Spent Mushroom Compost (SMC) + Sand + Lime (4:1:1)	83.34	0.250	4.46
CD(5%)			0.181

\*Average of 7 replications, \*\*Bag- 7 kg compost

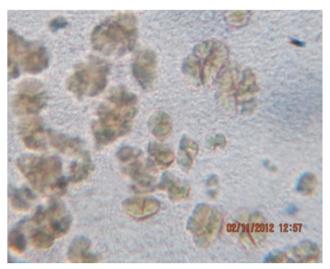


Figure 3: Microscopical examination of bulbils body of *Papulaspora byssina* (40x)

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### CONTENTS

		Page
<b>A.</b> 1.	<b>RESEARCH PAPER</b> The prevalence and etiology of polycystic ovarian syndrome (PCOS) as a cause of female infertility in central Travancore K. Roy George and N. A. Malini	- 001 - 006
2.	Diveristy analysis of different accessions of <i>Aloe barbadensis</i> Mill. (Syn. <i>Aloe vera</i> L.) collected from Rajasthan using RAPD marker system Dinesh Chandra and Preeti Choudhary	- 007 - 010
3.	In-vitro effects of herbicides on soil microbial communities	- 011 - 016
4.	Biology of <i>Coccinella transversalis</i> (Fabricius) on different aphid species Abhishek Shukla and Darshana S. Jadhav ————————————————————————————————————	- 017 - 022
5.	Effectiveness of synthetic insecticides against <i>Helicoverpa armigera</i> (Hubner) Hardwick and <i>Spodoptera litura</i> (Fabricius) infesting groundnut H. A. Gadhiya, P. K. Borad and J. B. Bhut	- 023 - 026
6.	Effect of PGPR and organic manures on soil properties of organically cultivated mungbean Ipsita Das and A. P. Singh	- 027 - 029
7.	Productivity of maize (Zea mays) based intercropping system during kharif season under red and lateritic tract of West Bengal M. K. Mandal, M. Banerjee, H. Banerjee, A. Alipatra and G. C. Malik	- 031 - 035
8.	Effect of foliar application of growth retardant on yield and germinability of hybrid rice M. Thoithoi Devi, Omvati Verma, Maya Krishna and Seema	- 037 - 039
9.	Influence of plant densities and planting dates on the population of pigeonpea flower blister beetles in Owerri, Imo state, Nigeria S. A. Dialoke, F. O. Ojiako, B. O. Bosah and C. A. Peter-Onoh	- 041 - 044
10.	Effect of humic acid on plant growth characters and grain yield of drip fertigated aerobic rice ( <i>Oryza sativa</i> L.) K. Vanitha and S. Mohandass —	- 045 - 050
11.	Impact of shoot pruning on root distribution pattern of litchi ( <i>Litchi chinensis</i> Sonn.) Bikash Das —	- 051 - 053
12.	Influence of sett size, seed rate and sett treatment on yield and quality of sugarcane Darpana Patel and Rinku Patel —	- 055 - 057
13.	A study on variation in biochemical aspects of different tree species with tolerance and performance index Meha Bora and Namita Joshi	- 059 - 063
14.	Temporal and intraday abundance variations of blister beetle ( <i>Mylabris phalerata</i> ) on greengram K. S. Pawar, S. P. Dhavan and R. M. Wadaskar —	- 065 - 069
15.	Effect of some botanicals on biological parameters of pulse beetle ( <i>Callosobruchus chinensis</i> L.) in pea ( <i>Pisum sativum</i> L.) Tarak Brambha Maji, S. Pal and H. Chatterjee	- 071 - 074
16.	Impact of honey bee pollination on pod set of mustard ( <i>Brassica juncea</i> L. : Cruciferae) at Pantnagar Vimla Goswami and M. S. Khan	- 075 - 078
17.	Effect of growing media properties and its correlation study in gerbera production F. G. Panj, Sunila Kumari and P. B. Parmar	- 079 - 083
18.	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	- 085 - 088
19.	Bio-efficacy of cyantrannilliprole 10% OD - An anthranilic diamide insecticide against sucking pests of cotton R. D. Patel, T. M. Bharpoda, N. B. Patel and P. K. Borad	- 089 <b>-</b> 092



		Pag
20.	Effect of sole and combined inoculation of <i>Ralstonia solanacearum</i> and <i>Meloidogyne javanica</i> on tomato P. P. Ghosh and S. Dutta	-093 - 100
21.	Integration approach in nutrient management of sesame with special reference to its yield, quality and nutrient uptake Snehangshu Sekhar Nayek, Koushik Brahmachari, Md. Riton Choudhury—	-101 - 105
22.	Productivity enhancement of sesame ( <i>Sesamum indicum</i> L.) through improved production technologies Rudrasen Singh, A. K. Upadhyay, Prashant Shrivastava, V. K. Singh and S. K. Singh	-107 - 110
23.	Effect of seed priming on field performance and seed yield of soybean [ <i>Glycine max</i> (L.) Merill] varieties N. G. Chavan, G. B. Bhujbal and M. R. Manjare	
24.	Effect of nitrogen and phosphorus on nutrient content and uptake in different varieties of African marigold ( <i>Tagetes erecta</i> L.) N. D. Polara, N. N. Gajpara and A. V. Barad	- 115 - 119
25.	Performance of growing calves fed on banana ( <i>Musa paradisiaca</i> ) stem Bhim Sen, Jai Singh, Tarun Verma and Prabhat R. Patel	
26.	Regeneration media standardization and molecular diversity analysis in cotton cultivars A. A. Bharose, S. A. Thakur and D. N. Damse ————————————————————————————————————	- 125 - 131
27.	Impact of bio-fertilizer seed treatment on seed and seedling parameters of maize (Zea mays L.) Animesh Pathak and S. K. Chakraborti	- 133 - 135
28.	Influence of sulphur and zinc fertilization on yield, yield components and quality traits of soybean ( <i>Glycine max</i> L.) Pratibha Choudhary, Arun Jhajharia and Rohit Kumar	
29.	Optimization of best cultural conditions for high production of phosphate solublizing activity by fluorescent <i>Pseudomonas</i> isolated from normal and replant sites of apple and pear Deepshikha Thakur, Mohinder Kaur and Vineet Shyam	
30.	Growth and available nutrient in winter maize ( <i>Zea mays</i> L.) with vegetable intercrops in eastern Uttar Pradesh S. K. Choudhary, R. N. Singh, P. K. Upadhyay, R. K. Singh and R. I. Yadav —	
81.	Effect of chemicals and growth regulators on germination, vigour and growth of passion fruit ( <i>Passiflora edulis</i> Sims.) N. Gurung, G. S. K. Swamy, S. K. Sarkar and N. B. Ubale	- 155 - 157
32.	Response of soil and foliar application of silicon and micro nutrients on leaf nutrient status of sapota K. A. Lalithya, H. P. Bhagya, K. Bharathi and Kulapati Hipparagi	
33.	Effect of nitrogen and zinc fertilizer on pearl millet ( <i>Pennisetum glaucum</i> ) under agri-horti system of eastern Uttar Pradesh S. K. Prasad, M. K. Singh and Renu Singh—	- 163 - 166
34.	Study of gonadosomatic index and fecundity of fish <i>Cirrhinus mrigala</i> (Hamilton) V. R. Chavan and D. V. Muley	- 167 - 169
35.	Response of soil and foliar application of silicon and micro nutrients on soil nutrient availability of sapota K. A. Lalithya, H. P. Bhagya, Amreen Taj, K. Bharati and Kulapati Hipparagi	- 171 - 174
36.	Evaluation of some fungicides, botanicals and essential oils against the fungus <i>Colletotrichum falcatum</i> causing red rot of sugarcane Nikhil Bhardwaj and R. K. Sahu —	
37.	Evaluation of bioagents for management of the onion purple blotch and bulb yield loss assessment under field conditions	- 179 - 181
88.	Bionomics and evaluation of different bio pesticides against <i>Helicoverpa armigera</i> (Hubner) Hardwick infesting groundnut H. A. Gadhiya, P. K. Borad and J. B. Bhut —	
39.	Management of vascular wilt of lentil through oils Vijay Kumar, Ankita Garkoti and H. S. Tripathi	- 189 - 192
10.	Yield loss assessment and influence of temperature and relative humidity on charcoal rot development in sesame (Sesamum indicum L)	
	P. Deepthi, C. S. Shukla, K. P. Verma and Siva Sankar Reddy E.	- 193 - 195
1.	P. Deepthi, C. S. Shukia, K. P. Verma and SiVa Sankar Reddy E. — Management of fusarium wilt of tomato by weeds and mycoflora processed weeds compost Vibha and Nidhi —	- 197 - 202
2.	Studies on foliar application of growth regulators and chemicals on seedling growth of mango varieties B. M. Muralidhara, Y. T. N. Reddy, M. K. Shivaprasad, H. J. Akshitha and Kishor Kumar Mahanthi	
13.	Diversity of orthopteran fauna in sugarcane at Udaipur Devendra Dhakad, Rajendra Nagar, Jhabar Mal, P. S. Rathore and R. Swaminathan	
14.	Serum IGE level in mice infected with single doses of <i>Ancylostoma caninum</i> larvae V. Viveka Vardhani and G. Sakunthala	
45.	Management of yellow rust ( <i>Pucciniastriiformis</i> . West) of wheat and its impact on yield under Jammu sub-tropics of India Rayees A. Ahanger, Vishal Gupta, Hilal A. Bhat and Nisar A. Dar	